



Global Transcriptome Analysis of Pseudomonas aeruginosa Response to Ortho-phenylphenol

48th Annual ICAAC/ IDSA 46th Annual Meeting

Background

- June 26, 2000 President Clinton met with the Director of the Human Genome Program and the CEO of Celera Genomics announcing the completion of the sequencing of the Human Genome
- August 2000, the complete genome sequence of *Pseudomonas aeruginosa* PA01 published (Cystic Fibrosis Foundation)
- July 2003 the Microarray Research Laboratory (MARL) was established at Fort Meade, MD

Pseudomonas aeruginosa: Nosocomial infections

- Nosocomial infections:
 - Estimated to occur in 5% of all acute-care hospitalizations.
 - More than 2 million cases each year
 - Cost of 4.5 billion dollars but most importantly 90,000 die.

• P. aeruginosa

- Gram negative rods
- Most common opportunistic pathogen
- Cystic fibrosis patients: Chronic lung infections



Increasing prevalence of nosocomial infections:
 Linked to increasing antimicrobial and disinfectant resistant pathogens.

Ortho-phenylphenol and P. aeruginosa

- Ortho-phenylphenol (OPP):
 - EPA approved chemical
 - Active ingredient in disinfectants.
 - Mode of action in bacteria has not been elucidated
- Use of OPP as a hospital disinfectant necessitates an understanding of the cellular functions that it affects in different pathogenic bacteria.
 - Facilitate determination of mode of action
 - Development of antimicrobials which target specific pathogenic bacteria and exert nominal effects on other species

GOALS

 What genes, proteins (enzymes), and ultimately metabolic pathways are affected in *P. aeruginosa* as a result of OPP treatment?

• What are the potential modes of action by which OPP inhibits *P. aeruginosa* growth?

Methods

- Sublethal concentration of OPP that will produce strong growth inhibition: <u>0.82mM</u>
- Early and late transcriptomic response to OPP : RNA extracted after <u>20</u> and <u>60</u> minutes.



Agilent 2100 Bioanalyzer & RNA LabChip







Methods

- 4 replicates each: control, 20 min, 60min
- P. aeruginosa GeneChip arrays (Affymetrix)
- Real-time PCR: Validation of microarray results



Analysis and Results

- GeneChip Operating Software (Affymetrix)
- GeneSpring Software(Agilent Technologies)
- One-way ANOVA: 1012 out of 5900 genes (*P. aeruginosa* genome) were statistically significant (p≤0.05).
- Fold Changes: Calculated as the ratios between the signal averages of four untreated (control) and four OPP-treated (experimental) cultures.
 - 509 genes: Upregulated (≥ 2-fold) and downregulated (≤ 2-fold) after 20 and 60 minutes exposure to 0.82mMOPP

- Functional classes: P. aeruginosa Community Annotation Project
- Upregulation: Filled bars
- Downregulation: Empty bars



Group I: Up 20, Up 60Group II: Up 20, No change 60Group III: Down 20, No change 60

Group IV: No change 20, Up 60 Group V: No change 20, Down 60

Group VI: Down 20, Down 60



Number of genes

Functional Classes

- Group I : Up 20 min; Up 60 min
 - Genes encoding 30 and 50s ribosomal proteins, translation initiation and elongation factors.
 - Membrane transport proteins: secY, secE and secG.
 - Virulence genes: *hit*A (ferric iron binding periplasmic protein); *hit*B (Iron III transport system permease).
 - Type IV pilus assembly proteins: *pil* C, D, G, I, M, N, O and P.
 - Upregulation of virulence genes => protective response to OPP treatment.

- Group II : Up 20 min; No change 60 min
 - norB- nitric oxide reductase subunit B: 4-fold upregulation.
 - Nitric oxide reductase enzyme :expressed under anaerobic conditions in *P. stutzeri*.
 - Possible shift to anaerobic respiration after 20 min : Nitrate used as final electron acceptor – Denitrification.
 - Rhamnosyl transferase chain A (rhlA): 2.4 fold upregulation
 - *rhl*A: critical for the exhibition of swarming motility by *P.* aeruginosa – Environmental adaptation.

- Group III : Down 20 min; No change 60 min
 - hcnA, hcnB, hcnC: Approximately 2-fold downregulation.
 - hcnABC encodes a cyanide synthase, which forms hydrogen cyanide from glycine.
 - *P. aeruginosa* does not produce cyanide under anaerobic conditions: nitrate being used as the terminal electron acceptor.
 - Supports theory: Possible transient switch to anaerobic respiration after 20 minutes of OPP treatment.

Results and Discussion Group IV : No change 20 min; Up 60 min

- Amino acid biosynthesis genes: argG, argH, glnA, lysA, lysC, proA, gltP, hisB, hisE, aroK, serC, glyA3.
- Contrasting results: In S. aureus exposed to OPP, amino acid biosynthesis genes are downregulated.
- Specifically lysine and diaminopimelic acid (DAP) biosynthesis were markedly downregulated : Inhibition of peptidoglycan layer formation =>Possible mode of action of OPP on S. aureus.
- This suggests that the effect of 0.82mM OPP in *P. aeruginosa* and *S. aureus* differ.
- Implications for Disinfectant choices for eliminating different bacteria in hospitals.

- Group V : No change 20 min; Down 60 min
 - napA, B, D and F genes: Components of the nap operon that encodes a periplasmic nitrate reductase.
 - The periplasmic nitrate reductase supports anaerobic growth in the presence of nitrate : Denitrification.
 - Implication: Anaerobic respiration is not favored after 60 minutes of OPP exposure
 - Contrast to after 20 minutes when anaerobic growth is favored.

- Group VI : Down 20 min; Down 60 min
 - Most downregulated gene: ribosome modulation factor (*rmf*)
 Fold change: -6.25 after 20 minutes and -25.9 after 60 minutes.
 - RMF: promotes efficient protein synthesis
 - The rpoS gene: Encodes RpoS, an alternative sigma factor of RNA ploymerase.
 - RpoS: Master regulator of gene expression in exponentially growing *E. coli* cells exposed to osmotic stress.
 - In *E. coli*: Mutations in *rmf* and *rpoS* => Decreases in cell viability.
 - Downregulation of *rmf* and *rpoS* may be related to the mechanism by which OPP causes growth inhibition in *P.* aeruginosa.

Conclusions

- When exposed to 0.82mM OPP, *P. aeruginosa* may switch to anaerobic respiration after 20 minutes and resume aerobic respiration after 60 minutes.
- Downregulation of *rmf* and *rpoS* may be related to the mechanism by which OPP causes growth inhibition in *P. aeruginosa*.
- Response to OPP exposure includes the upregulation of translation leading to the production of membrane transport and virulence proteins.
- Effect of OPP on *P. aeruginosa* and *S. aureus* differ.
- This gene expression profile can be used for a better understanding of:
 - The target cellular pathways of OPP in *P. aeruginosa*.
 - How *P. aeruginosa* develops resistance to OPP.

Chantal W. Nde, Hyeung-Jin Jang, Freshteh Toghrol and William E. Bentley

Lab Website:http://marl.umd.edu/index.html

University of Maryland & United States Environmental Protection Agency